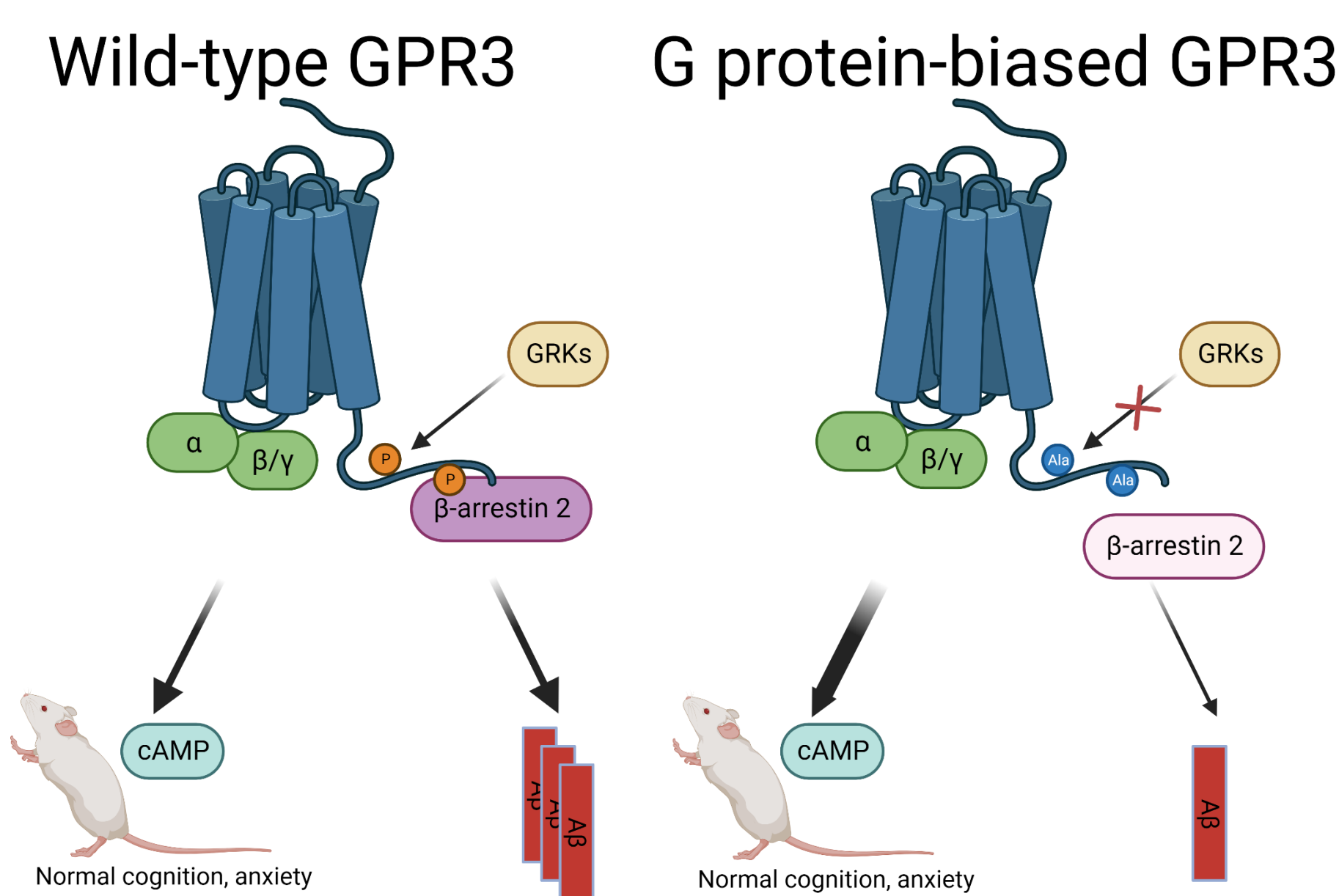


BACKGROUND

- Alzheimer's Disease (AD) is a neurodegenerative disorder pathologically characterized by the accumulation of intracellular neuronal tau tangles and extracellular amyloid-beta (Aβ) plaques¹.
- Biased G protein-coupled receptor (GPCR) signaling preferentially activates G protein- or β-arrestin-mediated signaling pathways and presents opportunities to develop more selective and safer therapeutics in the absence of side effects².
- Although GPCRs are implicated in the pathophysiology of AD³, biased GPCR signaling is a largely unexplored area of investigation in AD.
- Recently, we developed a G protein-biased GPR3 AD mouse model, which does not recruit β-arrestin 2, that displays reduced Aβ pathology without adverse cognitive effects⁴. the



OBJECTIVE

To investigate the mechanism by which G protein-biased GPR3 signaling ameliorates Aβ pathology using *in vitro* cellular models

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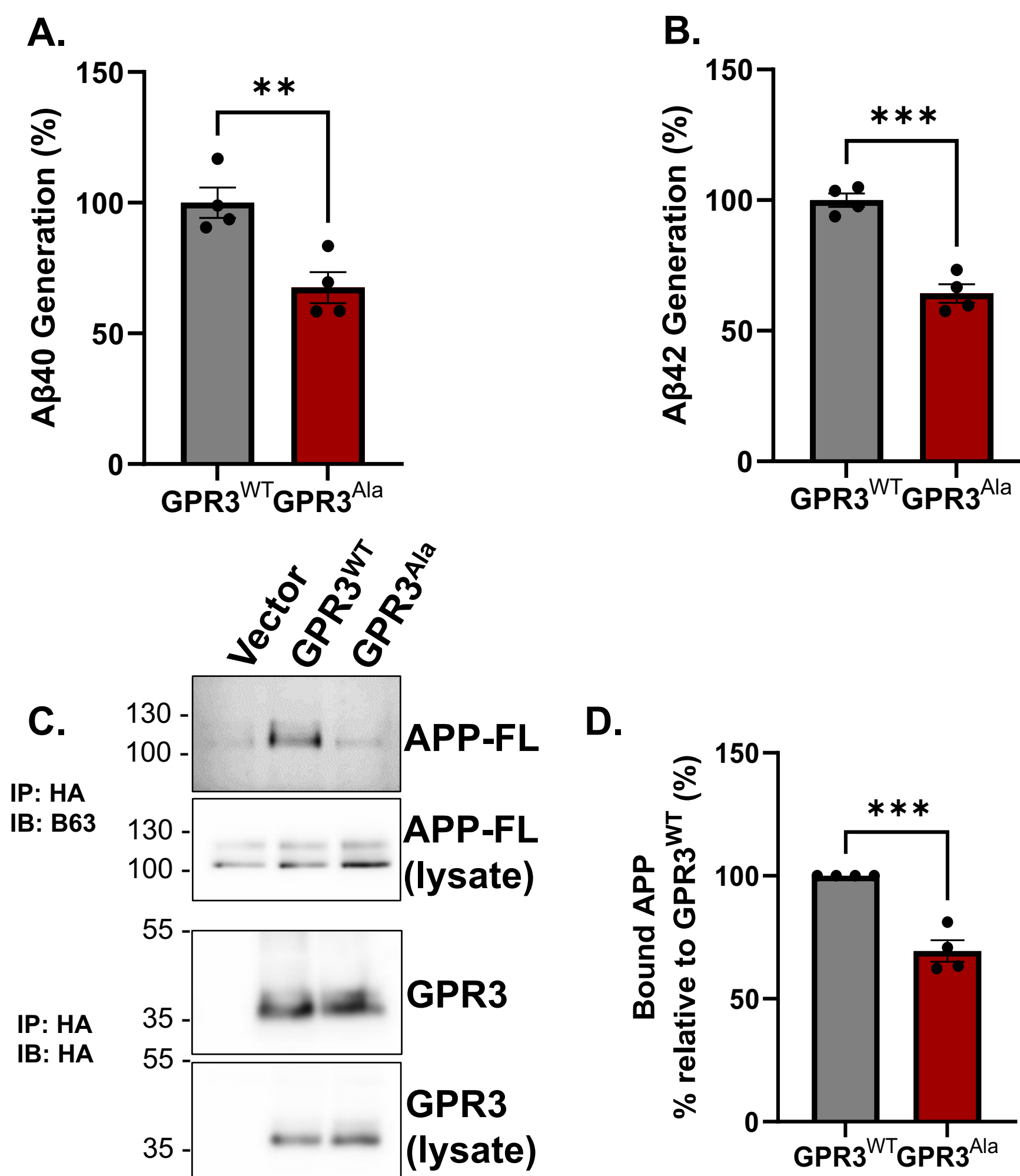
ACKNOWLEDGMENTS

The support and guidance from all past and present Thatthiah lab members has been indispensable for the completion of this work. We greatly appreciate the guidance and support of our collaborators and their laboratory members for their indispensable help with sharing of materials and methodological consulting.

Sources of support: NIA R01 AG058851 and Clear Thoughts Foundation grant.

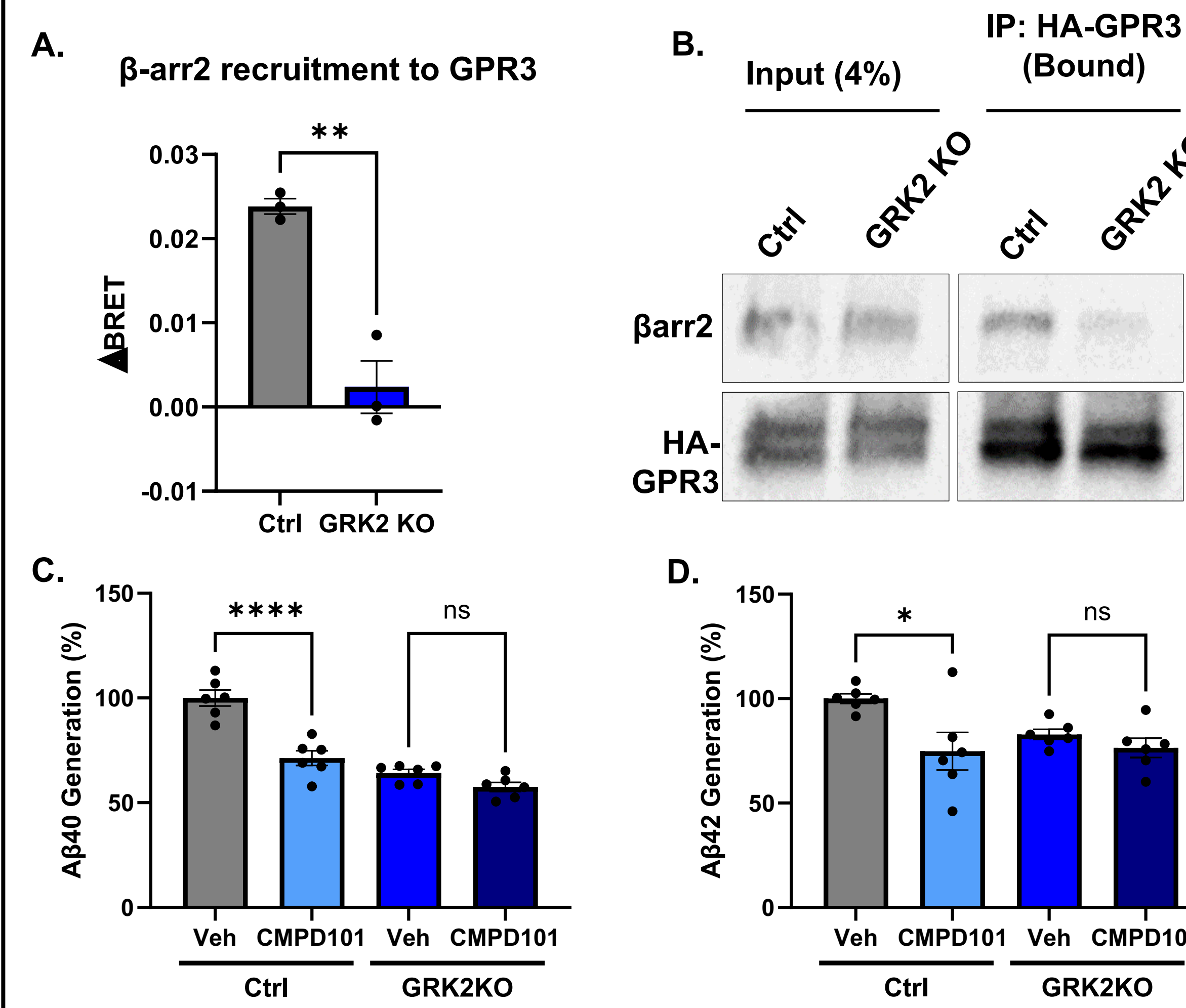
RESULTS

Figure 1. G protein-biased GPR3 displays reduced amyloid precursor protein (APP) binding and Aβ generation



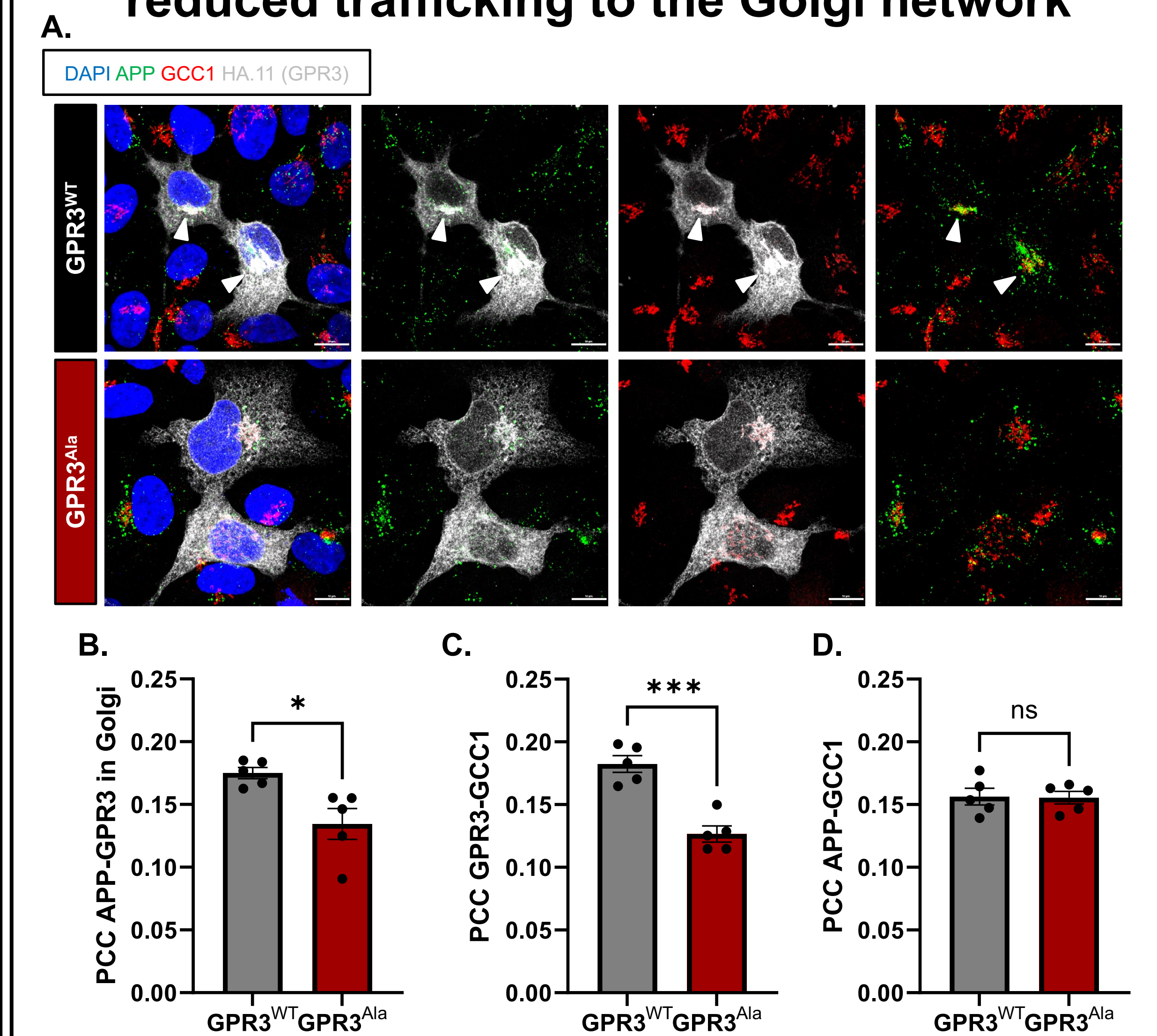
(A and B) Aβ40 (A) and Aβ42 (B) generation in HEK293 cells stably expressing APP695 (HEK-APP) and transfected with wild-type GPR3 (GPR3^{WT}) or G protein-biased GPR3 (GPR3^{Ala}). Data as mean±SEM. n=4 independent experiments. Unpaired two-tailed t-test. ** = p<0.01. *** = p<0.001. (C and D) Representative co-immunoprecipitation (C) and quantification (D) of full-length APP and GPR3^{WT} or GPR3^{Ala} from HEK-APP lysates. Data as mean±SEM. n=4 independent experiments. Unpaired two-tailed t-test. *** = p<0.001. Molecular weight in kDa.

Figure 2. GRK2 is involved in β-arrestin 2 recruitment to GPR3 and Aβ generation



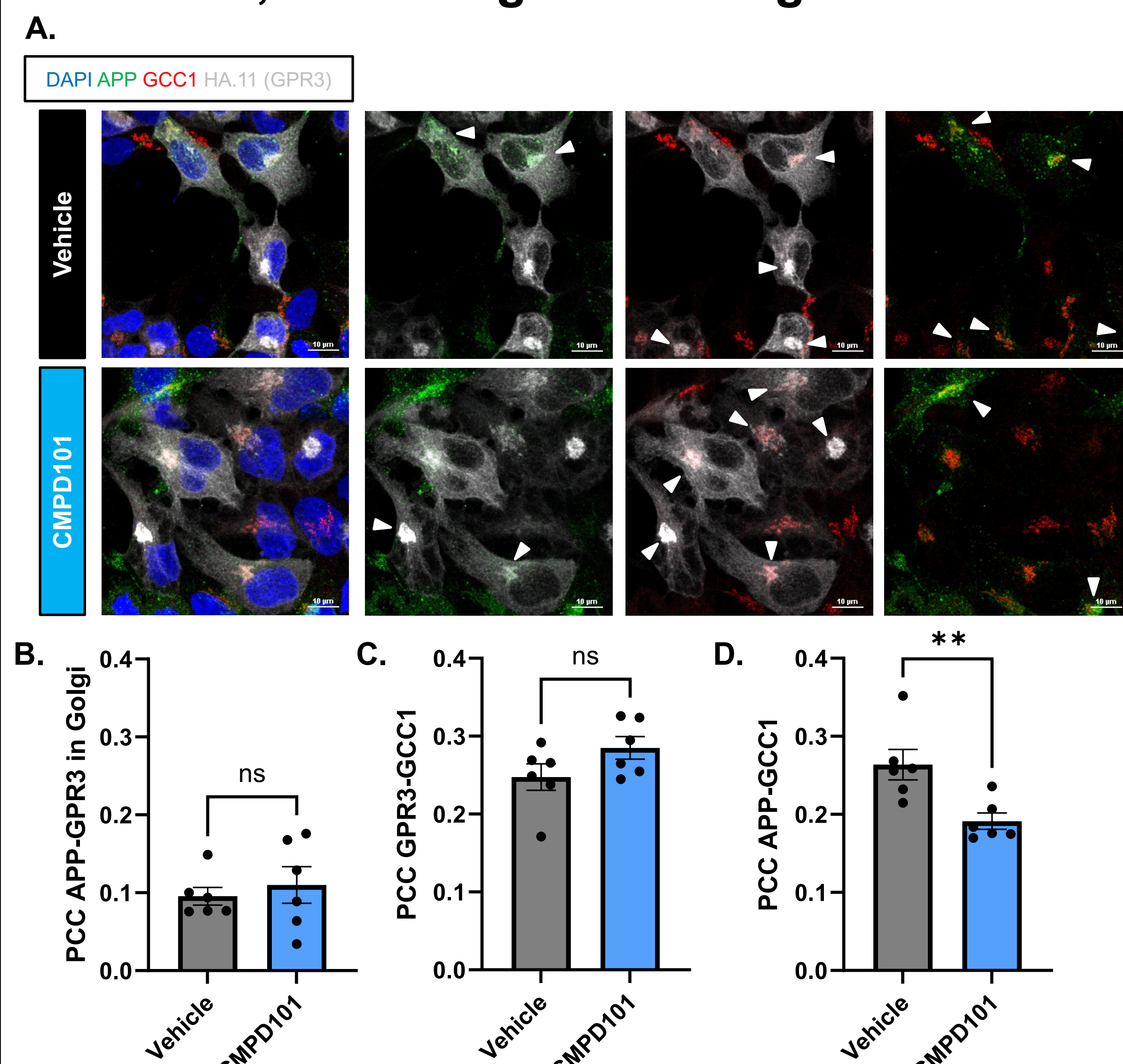
(A) BRET assay measuring βarr2 recruitment to GPR3^{WT} in HEK293 control and GRK2 KO cells. Data as mean±SEM. n=3 independent experiments. Unpaired two-tailed t-test. ** = p<0.01 (B) Co-immunoprecipitation experiment in HEK293 control or GRK2 KO cells expressing GPR3^{WT}. (C and D) Aβ40 (C) and Aβ42 (D) generation in HEK293 CRISPR control and GRK2 KO cells expressing APP-C99 and GPR3^{WT} ± 10 μM CMPD101. Data as mean±SEM. n=3 independent experiments run with 2 technical replicates each. Ordinary two-way ANOVA with Tukey's multiple comparisons test. * = p<0.05. **** = p<0.0001. ns = not significant.

Figure 3. G protein-biased GPR3 displays reduced trafficking to the Golgi network



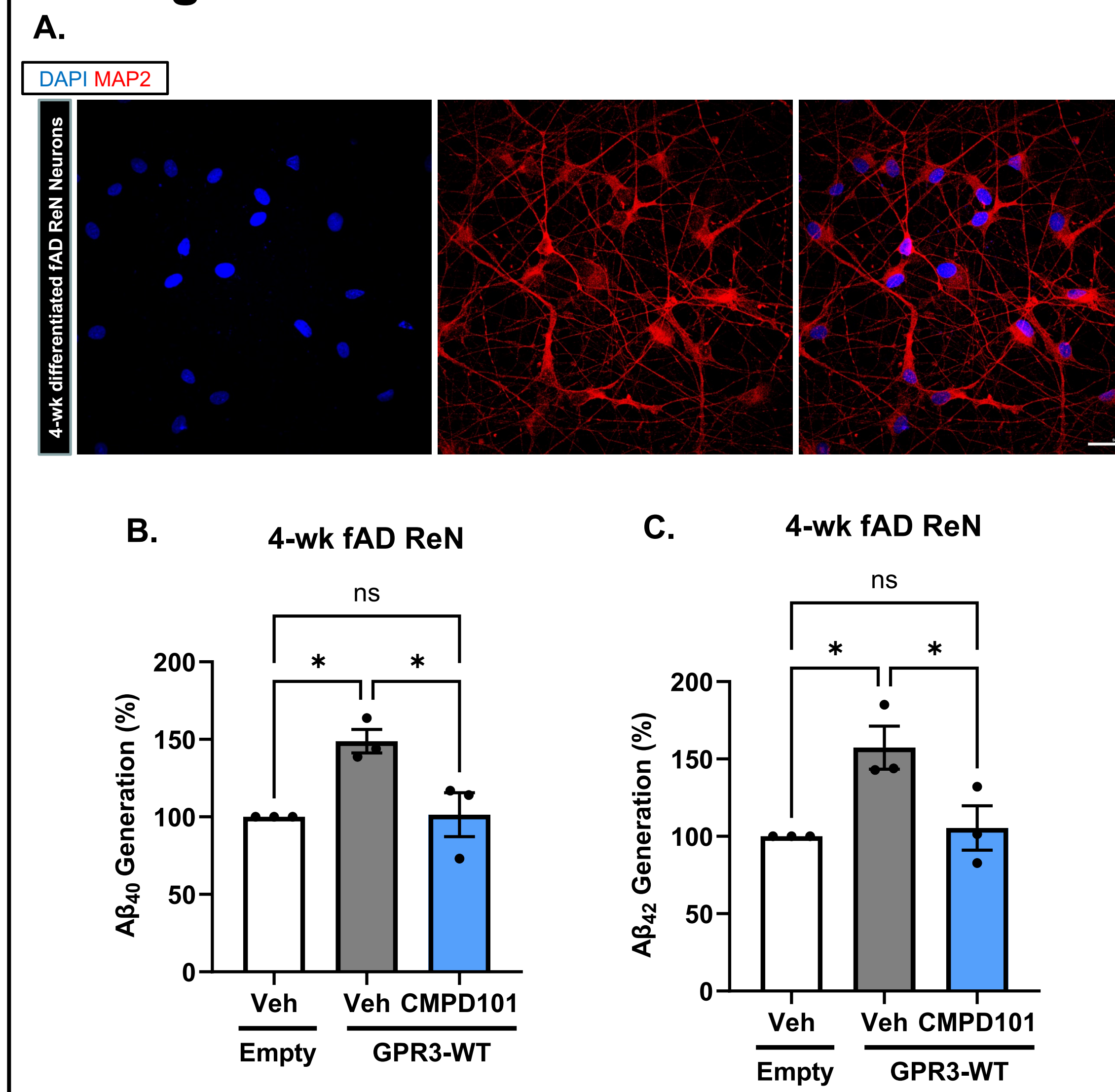
(A) Representative confocal images of HEK-APP cells expressing GPR3^{WT} or GPR3^{Ala}. Scale bars are 10 μm. (B-D) Quantification of colocalization of APP and GPR3 in Golgi binary (B), GPR3 and GCC1 (C), and APP and GCC1 (D) with Pearson correlation coefficient. Data as mean±SEM. n=5 independent experiments, two technical replicates each. Unpaired two-tailed t-test. * = p<0.05. *** = p<0.001. ns = not significant.

Figure 4. GRK2 activity affects APP, but not GPR3, trafficking to the Golgi network



(A) Representative confocal images of HEK-APP cells expressing GPR3^{WT} ± 100 μM CMPD101. Scale bars are 10 μm. (B-D) Quantification of colocalization of APP and GPR3 in Golgi (B), GPR3 and GCC1 (C), and APP and GCC1 (D) with Pearson correlation coefficient. Data as mean±SEM. n=6 independent experiments. Paired two-tailed t-test. ** = p<0.01. ns = not significant.

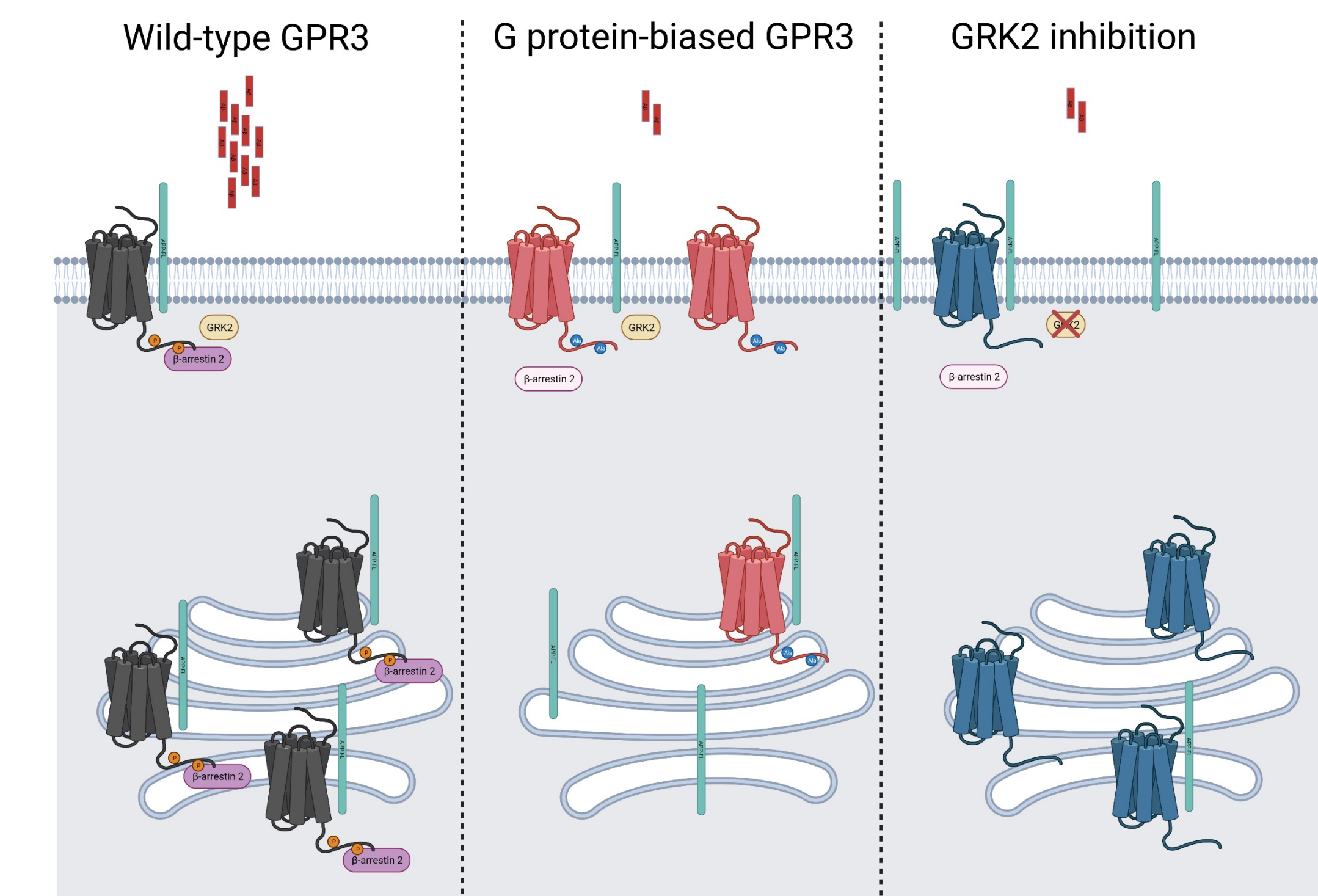
Figure 5. GRK2 inhibition reduces Aβ generation in human fAD neurons



(A) Representative confocal images of 4-week differentiated fAD neurons. Scale bars are 50 μm. (B and C) Aβ40 (B) and Aβ42 (C) generation in 4-week differentiated fAD neurons with and without GPR3^{WT} expression and ± 10 μM CMPD101. Data as mean±SEM. n=3 independent experiments. Ordinary one-way ANOVA with Tukey's multiple comparisons test. * = p<0.05. ns = not significant.

SUMMARY

- G protein-biased GPR3 shows reduced binding to APP and reduced localization to the Golgi network.
- GRK2 is involved in β-arrestin 2 recruitment to GPR3 and Aβ generation.
- Inhibition of GRK2 leads to a reduction in APP localization to the Golgi network.



NEXT STEPS

- Identify putative GRK phosphorylation sites on GPR3 and APP
- Determine the effect of GRK2 inhibition on GPR3-APP binding
- Investigate the impact of biased GPR3 or GRK2 inhibition on GPR3 and APP trafficking to membrane/other subcellular compartments